STRUCTURE OF INFLEXINOL, A NEW CYTOTOXIC DITERPENE FROM RABDOSIA INFLEXA

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Abstract—From the leaves of Rabdosia inflexa a new cytotoxic diterpenoid, inflexinol was isolated, together with the known inflexin, and the structure was established as ent-kaur-16-en-15-one- 1α , 3α , 6β , 11β -tetraol 1.3-diacetate.

INTRODUCTION

From the various plants, belonging to the genus Rabdosia [1] (Labiatae), many diterpenoids of the ent-kaurene, B-seco-ent-kaurene and B/C-seco-ent-kaurene types were isolated and their structures were determined [2, 3]. Recently, antitumor and antibacterial activities of these diterpenoids were examined [4, 5]. During the investigations of biologically active substances, we have examined the constituents of the leaves of Rabdosia inflexa and isolated a new minor diterpenoid, inflexinol (1) together with the known inflexin (2) [6]. Inflexinol (1) showed a cytotoxic activity against cultured rat mammary cancer FM 3A/B cells. We report here the structure elucidation of inflexinol (1).

RESULTS AND DISCUSSION

Inflexinol (1) was obtained from the methanolic extract of the leaves of R. inflexa as an amorphous powder, $[\alpha]_D^{28} - 43.0^{\circ}$ (MeOH; c 0.22) and had a molecular formula, C₂₄H₃₄O₇, from high resolution mass spectroscopy. Inflexinol (1) showed an absorption maximum at 237 nm (ϵ 6124) in the UV, absorptions at 1720 and 1645 cm⁻¹ in the IR and signals at δ 5.29 and 5.90 (each 1 H, each br s) in the ¹HNMR spectrum. These spectral data suggested that 1 had a five membered ketone conjugated with an α -methylene group as a partial structure. Besides the signals of three tertiary methyl groups at δ 0.91, 1.28 and 1.49 and the signals of two acetyl groups at 1.90 and 2.07, the 'H NMR spectrum showed the signals due to two protons attached to carbons having an acetoxy group at 4.71 (t, J = 3 Hz, H_b) and 5.90 (t, J = 3 Hz, H_a) and the signals assigned to protons attached to a hydroxy group bearing carbons at 3.69 $(dd, J = 10 \text{ and } 6 \text{ Hz}, H_d) \text{ and } 4.43 (dd, J = 5 \text{ and } 4.43)$ 3 Hz, H_c). From these facts, it was presumed that inflexinol (1) had a 15-oxo-ent-kaurene as a basic skeleton which is common in the diterpenoids of Rabdosia plants. In fact, dihydroinflexinol (3) showed

a negative Cotton effect in its ORD spectrum. Comparing the ¹H NMR spectrum of 1 with that of 2, the proton signals due to H_a , H_b and H_d in 1 were observed at the corresponding position in 2, but the new proton signal due to H_c also appeared in the spectrum of 1. On the other hand, the signals at δ 2.71 (s, 5-H) and 3.12 (d, J = 12 Hz, 7α -H) in the spectrum of 2 were missing in that of 1. These results suggest that the carbonyl group at C-6 in 2 had been reduced to an alcoholic group in 1. We have further examined the probability of this presumption by INDOR (internuclear double resonance) [7] experiments.

On monitoring H_g or H_i, the INDOR signals were observed at H_e and H_i, or H_e and H_g. These results support the structure of the D-ring. When H_b was monitored, an INDOR signal due to coupling was observed in the same methylene region (δ 1.60–2.40) as on monitoring H_a and the signals due to NOE's were also observed on both the methyl groups at C-4 $(\delta 0.91 \text{ and } 1.28)$. These results suggested that 1 had β-axial acetoxy groups at C-1 and C-3 in the A-ring as in the case of 2. On monitoring H_c, INDOR signals due to coupling were observed around δ 1.29-1.42 and 2.00-2.29 and the signals due to NOE's were observed for methyl groups at C-4. Considering the coupling constant of H_c, this result suggested that H_c was a β -equatorial proton at C-6, namely, an α -axial hydroxy group was located at that position. Finally, a signal arising from NOE was observed on H_f (δ 2.91, d, J = 13 Hz, 14α -H) when monitored from a methyl group at C-10 (δ 1.49). From the above-mentioned facts, the structure of inflexinol must be ent-kaur-16en-15-one- 1α , 3α , 6β , 11β -tetraol 1, 3-diacetate (1).

In order to verify the presumed structure, inflexinol (1) was chemically correlated with inflexin (2). Acetylation of 1 with Ac_2O -pyridine gave 11-monoacetate (4), mp 247-248° (δ 4.84, dd, J = 12 and 6 Hz, 11 β -H). The fact that a hydroxy group at C-6 was not acetylated agrees with the presumption that the configuration of the hydroxy group is α -axial.

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Jones oxidation of 4 gave inflexin acetate (5). When dihydroinflexinol (3) was subjected to LiAlH₄ reduction, a pentahydroxy-ent-kaurane (6), mp 227-229°, was obtained. From the consideration of the stereomodel, it is expected that 6α -axial hydroxy group should be formed by LiAlH₄ reduction of dihydroinflexin (7) as in the case of 6-ketosteroids [8]. In fact, the product obtained from 7 was identical with 6. Accordingly, the structure of inflexinol was established as 1.

EXPERIMENTAL

General procedures. All mps are uncorr. ¹H NMR: 100 or 90 MHz. All the ¹H NMR spectra were taken for CDCl₃ solns unless otherwise noted. Chemical shifts are given in δ with TMS as int. standard. MS: 70 eV unless otherwise noted.

Plant material. Plants were collected in the suburbs of Tokushima City (Tokushima Pref., Japan) in Oct. 1976 and identified as Rabdosia inflexa (Thunb.) Hara by Mr. G. Murata of Faculty of Sciences, Kyoto University. A voucher specimen (T. Fujita No. 12) was deposited in the Herbarium of the Institute of Botany, Kyoto University (KYO), Kitashirakawaoiwake-cho, Sakyo-ku, Kyoto 606, Japan.

Isolation procedures. Methanolic extracts obtained from the dried leaves of Rabdosia inflexa (6.2 kg) were combined and concd in vacuo. The residue was dissolved in 90% MeOH (31.) and then the soln was washed $\times 3$ with n-hexane (total 4.51.). The 90% MeOH layer was concd in vacuo. The residue was suspended in H_2O (1.61.) and extracted with EtOAc (total 7.51.). After washing with H_2O , the EtOAc extract was dried and evaporated in vacuo to give a residue (180.6 g). This residue was chromatographed on a Si gel (3 kg) column with CHCl₃-MeOH with increasing MeOH content. The fraction eluted with CHCl₃-MeOH (97:3) gave a residue (49.11 g), which contained inflexinol (1) and inflexin (2). The residue was further separated and purified by repeated Si gel chromatography using Et₂O as

eluent to give inflexinol (1) (0.435 g) and inflexin (2) (7.72 g). Inflexinol (1); IR $\nu_{\max}^{\text{CHCl}_3}$ cm⁻¹: 3600, 3550-3350, 1720, 1645, 1260; ¹HNMR: δ 0.91, 1.28, 1.49 (3 × s, 3 × tert. Me), 1.90, 2.07 (2 × s, 2 × OAc), 2.91 (d, J = 13 Hz, 14α -H), 3.11 (m, 13-H), 3.69 (dd, J = 12 and 5 Hz, 11-H), 4.43 (dd, J = 5 and 3 Hz, 6-H), 4.71 (t, J = 3 Hz, 3-H), 5.29 (br s, 17-H₁), 5.90 (t, J = 3 Hz, 1-H), 5.90 (br s, 17-H₁); MS m/z: M⁺ 434.230 (calc. for $C_{24}H_{34}O_7$ 434.230). Inflexinol (1) at 10 μ g/ml showed inhibitory activity (89.5%) on the growth of cultured rat mammary cancer FM 3A/B cells.

Inflexin (2); recrystallized from a mixture of Et₂O and *n*-hexane, mp 202-204°, $[\alpha]_D^{29} - 57.1°$ (MeOH; *c* 1.0); UV $\lambda_{\text{max}}^{\text{MOH}}$ nm (ϵ): 238 (7708); IR $\nu_{\text{max}}^{\text{MBr}}$ cm⁻¹: 3375, 1730, 1705, 1650, 1260; ¹H NMR: δ 0.86, 1.16, 1.32 (3 × *s*, 3 × tert. Me), 1.96, 2.12 (2 × *s*, 2 × OAc), 2.32 (*d*, J = 12 Hz, 14 α -H), 2.71 (*s*, 5-H), 3.10 (*m*, 13-H), 3.12 (*d*, J = 12 Hz, 7 α -H), 3.94 (*dd*, J = 12 and 6 Hz, 11-H), 4.56 (*t*, J = 3 Hz, 3-H), 5.34 (*br s*, 17-H₁), 5.95 (*t*, J = 3 Hz, 1-H), 5.95 (*br s*, 17-H₁); MS m/z: 432.217 [M]⁺ (calc. for C₂₄H₃₂O₇ 432.215). The physical properties of this substance agree with those of inflexin (2) [6].

Catalytic hydrogenation of inflexinol (1). Inflexinol (1) (85 mg) was dissolved in MeOH (8 ml) and hydrogenated over PtO₂ (4 mg) for 30 min. The catalyst was filtered off and the solvent removed in vacuo to give dihydroinflexinol (3) (74 mg). ORD $\lambda_{\text{max}}^{\text{MeOH}}$ nm ([ϕ]): 322 (-1831), 288 (+1716); IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3400, 2900, 1720, 1370, 1250; ¹H NMR: δ 0.91, 1.27, 1.48 (3 × s, 3 × tert. Me), 1.30 (d, J = 7 Hz, 16-Me), 1.99, 2.07 (2 × s, 2 × OAc), 2.88 (d, J = 12 Hz, 14 α -H), 3.64 (dd, J = 12 and 6 Hz, 11-H), 4.36 (m, 6-H), 4.67 (t, J = 3 Hz, 3-H), 5.76 (m, 1-H); MS (22 eV) m/z: 418.237 [M-H₂O]⁺ (calc. for $C_{24}H_{34}O_6$ 418.235).

Acetylation of inflexinol (1). Inflexinol (1) (50 mg) was acetylated with Ac_2O -pyridine at room temp. for 48 hr and the product purified on a Si gel column with CHCl₃ as eluent and recrystallized from a mixture of Et₂O and *n*-hexane to give inflexinol 11-acetate (4) (18 mg) as colourless plates, mp 247-248°. IR ν_{max}^{KBr} cm⁻¹: 3450, 2900, 1730, 1645, 1250; ¹H

NMR: δ 0.94, 1.30, 1.60 (3 × s, 3 × tert. Me), 1.78, 1.99, 2.09 (3 × s, 3 × OAc), 2.81 (d, J = 12 Hz, 14α -H), 3.06 (m, 13-H), 4.44 (m, 6-H), 4.64 (t, J = 3 Hz, 3-H), 4.85 (dd, J = 10 and 6 Hz, 11-H), 5.19 (br s, 17-H₁), 5.60 (m, 1-H), 5.84 (br s, 17-H₁) (Found C, 65.23; H, 7.88. $C_{26}H_{36}O_8$ requires: C, 65.53; H, 7.61%).

Acetylation of inflexin (2). Inflexin (2) (300 mg) was acetylated with Ac_2O -pyridine at room temp. for 84 hr to give inflexin acetate (5) (296 mg). $[\alpha]_D^{28} - 28.9^\circ$ (MeOH; c 0.27); IR $\nu_{\text{max}}^{\text{CCI}_4}$ cm⁻¹: 1740, 1720, 1645, 1370, 1245; ¹H NMR: δ 0.90, 1.25, 1.34 (3 × s, 3 × tert. Me), 1.84, 2.02, 2.13 (3 × s, 3 × OAc), 2.84 (s, 5-H), 3.06 (m, 13-H), 3.22 (d, J = 12 Hz, 7α -H), 4.57 (t, J = 3 Hz, 3-H), 5.08 (dd, J = 12 and 6 Hz, 11-H), 5.31 (br s, 17-H₁), 5.60 (m, 1-H), 5.95 (br s, 17-H₁); MS m/z: 474.224 [M]⁺ (calc. for $C_{26}H_{34}O_{8}$ 474.225).

Jones oxidation of inflexinol 11-acetate (4). A soln of 4 (18 mg) in Me₂CO (2 ml) was stirred with Jones reagent (5 drops) under ice cooling for 30 min. The reaction mixture was diluted with excess H₂O and extracted with CHCl₃. The CHCl₃ extract was dried and evaporated in vacuo. The residue (11 mg) was purified by Si gel TLC (CH₂Cl₂-Me₂CO, 9:1) to give an oxidation product (3.8 mg). $[\alpha]_{0}^{26} - 16.9^{\circ}$ (MeOH; c 0.21); MS m/z: 414.203 [M-AcOH]⁺ (calc. for C₂₄H₃₀O₆ 414.204). This substance was identified with an authentic sample of inflexin acetate (5) by comparison of IR and ¹H NMR spectra.

Reduction of dihydroinflexinol (3) with LiAlH₄. To a soln of dihydroinflexinol (3) (32 mg) in dry THF (10 ml), LiAlH₄ (32 mg) was added and the reaction mixture was stirred at room temp. for 24 hr. After successive addition of excess EtOAc, excess H₂O and dil. H₂SO₄ (until the resulting ppt. was dissolved), the resulting EtOAc phase was separated. The EtOAc extract was washed with H₂O, dried and evaporated in vacuo to give a residue (18 mg), which was purified by Si gel TLC (CHCl₃-MeOH, 9:1) and recrystallized from MeOH to give a pentahydroxy-ent-kaurane (6) (7.6 mg), mp 227-229°. IR $\nu_{\rm max}^{\rm Kgr}$ cm⁻¹: 3600-3000, 2860, 1060, 1030; ¹H NMR (d_5 -pyridine): δ 1.36, 1.64, 2.04 (3 × s, 3 × tert. Me), 1.46 (d, d = 8 Hz, 16-Me), 3.76 (1 H, d), 3.90 (1 H, d, d = 12 Hz), 4.72 (2 H, d), 5.08 (1 H, d); MS d = 336.234 [M-H₂O]⁺ (calc. for C₂₀H₃₂O₄ 336.230).

Catalytic hydrogenation of inflexin (2). Inflexin (2) (100 mg) was dissolved in MeOH (5 ml) and hydrogenated

over PtO₂ (5 mg) for 30 min. The catalyst was filtered off and solvent was removed in vacuo to give dihydroinflexin (7) (100 mg). IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 3250, 2850, 1730, 1715, 1365, 1230; $^{\rm 1}$ H NMR: δ 0.86, 1.12, 1.28 (3 × s, 3 × tert. Me), 1.29 (d, J=6 Hz, 16-Me), 2.01, 2.04 (2 × s, 2 × OAc), 2.34 (d, J=12 Hz, 14 α -H), 2.67 (s, 5-H), 3.02 (d, J=12 Hz, 7 α -H), 3.98 (dd, J=12 and 6 Hz, 11-H), 4.58 (t, J=3 Hz, 3-H), 5.80 (t, J=3 Hz, 1-H); MS m/z: 434.230 [M]⁺ (calc. for C₂₄H₃₄O₇ 434.230).

Reduction of dihydroinflexin (7) with LiAlH₄. To a soln of dihydroinflexin (7) (50 mg) in dry THF (5 ml), LiAlH₄ (50 mg) was added and the reaction mixture was stirred at room temp. for 4 hr. The reaction mixture was worked up as before and the product (20 mg) was recrystallized from MeOH to give pentahydroxy-ent-kaurene (6) (8 mg), mp 226-229°; MS m/z: 336.233 [M-H₂O]⁺ (calc. for C₂₀H₃₂O₄ 336.230). This substance was identified with 6, derived from dihydroinflexinol (3), by mmp and comparisons of IR and ¹H NMR spectra.

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REFERENCES

- 1. Hara, H. (1972) Jpn J. Botany 47, 193.
- 2. Fujita, E., Nagao, Y. and Node, M. (1976) Heterocycles 5, 793
- 3. Fujita, E., Ito, N., Uchida, I., Fuji, K., Taga, T. and Osaki, K. (1979) J. Chem. Soc. Chem. Commun. 806.
- 4. Fujita, E., Nagao, Y., Node, M., Kaneko, K., Nakazawa, S. and Kuroda, H. (1976) Experientia 32, 203.
- Yamaguchi, M., Taniguchi, M., Kubo, I. and Kubota, T. (1977) Agric. Biol. Chem. 41, 2475.
- Kubo, I., Nakanishi, K., Kamikawa, T., Isobe, T. and Kubota, T. (1977) Chem. Letters 99.
- Kikuchi, T., Yokoi, T., Niwa, M. and Shingu, T. (1980) Chem. Pharm. Bull. (Tokyo) 28, 2014.
- 8. Kirk, O. N. and Hartshorn, M. P. (1968) Steroid Reaction Mechanisms, p. 136. Elsevier, Amsterdam.